

High Risk Assessment of Pregnancy in Rasopathies

Ramachandran Muthiah*

President of all nations, Morning star hospital, Enayam Thoppu, Kanyakumari District, India

***Corresponding author:**

Ramachandran Muthiah, President of all Nations, Morning Star Hospital, Enayam Thoppu, Kanyakumari District, India.

Abstract

Rasopathies are the group of related disorders caused by abnormal functioning of the Ras-mitogen-activated protein kinase (RAS/MapK) pathway., a cohort of common congenital anomaly syndromes, occurring in 1:1,000 to 1:2,500 live births that have unique fetal features, which can be detected on routine prenatal ultrasounds. In the presence of suggestive US (ultrasound) findings, previous studies estimated that pathogenic variants in RASopathy genes could be detected in 6.7–21.7% of cases. Some disease-specific features, such as an increased prevalence of fetal arrhythmias among cases of Costello syndrome. Prenatal-onset HCM (hypertrophic cardiomyopathy) is rare and potentially severe to delineate high-risk genotype and so accurate diagnosis of fetal rasopathies is essential for improved management of affected pregnancies.

Keywords: Rasopathies, Fetal Anomalies, Ultrasound Features, High Risk in Pregnancy, Neonatal Complications, Parental Counselling.

Received: August 30, 2025;

Accepted: September 06, 2025;

Published: September 13, 2025

Introduction

RASopathies are a family of genetic disorders with more than 20 disease-associated genes are identified so far [1,2]. Many genetic syndromes, a part of RASopathy spectrum, including Noonan syndrome (NS), Noonan syndrome with multiple lentigines (previously known as LEOPARD syndrome), Noonan-like syndrome with loose anagen hair, Costello syndrome (CS), cardiofaciocutaneous syndrome (CFCS), and other clinically related disorders are frequently encountered in pre- and postnatal evaluations [3]. Prenatal diagnosis of RASopathies is important as it can improve parental counseling and allow families to make informed decisions with regard to pregnancy management, treatment options, living with a child with a genetic disorder, termination of pregnancy (TOP) and to screen for complications known to occur in pregnancies with RASopathies, like HCM, and prepare the medical team for management of neonatal complications, but challenging, mostly because of their variable expressivity, as well as their nonspecific prenatal presentation due to heterogeneous cohorts with overlapping prenatal phenotypes [4-6].

Etiopathogenesis

Researcher Susan White, a medical geneti-

cist at Royal Children's Hospital. Melbourne, Australia had a clinical and research interest in syndromes of childhood, worked with families for their child's suspected having genetic problems along with VCGS (Victorian Clinical Genetics Services) laboratory team and led the implementation of exome sequencing (a genomic technique for sequencing all of the protein-coding regions of genes in a genome known as the exome) [7].

Prenatal diagnosis with G-banded karyotyping to detect chromosomal abnormalities results in a diagnosis in 9 to 19% of fetal anomalies, and chromosomal microarray analysis provides an additional 6% yield. The cause of the majority of fetal anomalies remains unknown and Exome sequencing has transformed genetic diagnosis after birth and recent studies showed the diagnostic yields of 8.5% and 10% to 29 % in one series with rasopathies, but its usefulness in prenatal diagnosis is still emerging and Trio exome sequencing covers a wide spectrum of genetic change [8, 9]. In utero manifestations of RASopathies are less well characterized since approximately half of RASopathy variants being inherited in postnatal series and the de novo nature of all the RASopathy variants are severe with a wide range of outcomes, from

Citation: Ramachandran Muthiah (2025) High Risk Assessment of Pregnancy in Rasopathies. J Gyne Womens Heal Care 1: 1-11.

relatively mild lymphedema to probable perinatal death, and their clinical management differs greatly [10].

HRAS is a gene that provides instructions for making a protein called H-Ras which is involved primarily in regulating cell division and it is an enzyme in the signaling pathway (MAP kinase signaling cascade) that relays chemical signals from the outside of the cell to the cell's nucleus, telling the cell when to grow and divide and thus acts as a key regulator of MAPK (mitogen-activated protein kinase) pathway mediated by protein kinase Raf which is critical for cell cycle regulation, cellular differentiation and growth [11]. The HRAS gene is in the Ras family of oncogenes, which also includes two other genes, the KRAS and NRAS and the proteins produced from these three genes are GTPases (Guanosine triphosphate hydrolase), which means it converts a molecule called GTP into another molecule called GDP. The H-Ras protein acts like a switch as it is turned on by attaching (binding) to a molecule of GTP and turned off (inactivated) when it converts GTP to GDP [12]. After the protein is bound to GDP, it does not relay signals to the cell's nucleus. When mutated, HRAS can act as an oncogene, causing normal cells to become cancerous and perturbances (dysfunction) of this MAPK pathway during essential cellular functions and development leading to multiple consequences in embryonal and later stages of development [13].

A mutation is a change in a DNA sequence. DNA is a chain of many smaller molecules called nucleotides. During protein formation, DNA (or RNA) nucleotide sequences are read three nucleotides at a time in units called codons, and each codon corresponds to a specific amino acid or stop signal (stop codon). Stop codons are also called nonsense codons because they do not code for an amino acid and instead signal the end of protein synthesis [14]. Somatic mutations occur in a single body cell, cannot be inherited and only tissues derived from mutated cell are affected. Germline mutations occur in gametes, can be passed onto offspring and every cell in the entire organism will be affected. In contrast to somatic oncogenic mutations in neoplasia, the Costello syndrome changes are typically introduced in the paternal germline. Missense mutation is a genetic change that results in the substitution of one amino acid in protein for another [15]. A nonsense mutation is a genetic mutation in a DNA sequence that results in a shorter, unfinished protein product. Genetic heterogeneity can be defined as mutations at two or more genetic loci that produce the same or similar phenotypes (either biochemical or clinical) and it is relevant since it can present problems for heterozygote detection [16]. De novo mutation is a genetic alteration that is present for the first time in one family member as a result of a variant (or mutation) in a germ cell (egg or sperm) of one of the parents, or a variant that arises in the fertilized egg itself during early embryogenesis [17]. Codon encoding residues stabilizing the Ras protein in an inhibited conformation and SOS (son of sevenless)1 missense mutations disrupts the autoinhibition of Ras GEF (Guanine nucleotide exchange factor) activity and Ras GEF stimulate the conversion of Ras from inactive GDP bound form to active GTP- bound form. SOS1 gain-of-function increases the active form of Ras and thus increase the Ras/MAPK signaling. SOS-RAS activation is operated via FRS2 (Fibroblast growth factor receptor substrate 2) which acts downstream of TRKA (Tyrosine Kinase A) in neurons, and FGFR (Fibroblast growth factor receptor) in

embryonic stem cell [18]. HRAS (Harvey-Ras) was the first human proto-oncogene reported and p.Gly12Val in codon 12 is the first oncogenic mutation described and Costello syndrome was the first disorder associated with germline mutations in the RAS family of GTPases [19,20]. A nucleotide change that causes substitution of glycine at codon 12 to serine (p.G12S)) is the most common (80%), c.35G>C nucleotide resulting in p.Gly12Ala was seen in 9% and recently, p.Gly13Cys change was identified as most common amino acid change affecting the glycine in Position 13 [21,22]. Rare phenotypes often lethal in infancy are p.G12D, p.G12C, p.G12E and its manifestations are shown in Table 1 [23,24].

Table 1: Manifestations of Lethal Phenotypes in Infancy

S.no	Severe manifestations
1.	Hypoglycaemia
2.	renal abnormalities
3.	severe early cardiomyopathy
4.	congenital lung and airway abnormalities
5.	pleural and pericardial effusion
6.	chylous ascites
7.	pulmonary lymphangectasia
8.	alveolar capillary dysplasia.

Distinctive phenotypic findings of p.Gly13Cys (mild phenotype) are dolichocilia (extremely long eyelashes), loose anagen hair, fewer have short stature, no malignant risk, papillomata and multifocal atrial tachycardia are not seen [25]. HRAS mutations are associated with 10-15% of cancer risk. Germline HRAS mutations associated with Costello syndrome, which confers a risk for malignant tumors including rhabdomyosarcoma, neuroblastoma in childhood and bladder cancer in adolescence [26].

CBL (casitas B-cell lymphoma), a tumor suppressor gene encodes E3 ubiquitin ligase which negatively regulates Ras/MAPK signaling downstream of RTK (receptor tyrosine kinases) and mediates the association of ubiquitin with activated RTK, necessary for receptor internalization & degradation. Mutation in CBL reduce the turnover of activated RTK leads to increased ERK (Extracellular signal-regulated kinases) activation [27-29]. CBL is a cellular homolog of v-Cbl transforming gene of the Cas NS-1 murine leukemia virus, responsible for intracellular transport and degradation of a large number of proteins and majority of CBL somatic mutations are reported in myelodysplastic/myeloproliferative disorders such as chronic myelomonocytic, juvenile myelomonocytic, atypical chronic myeloid leukemias and germline mutations in CBL are identified in juvenile myelomonocytic leukemia [30]. Introduction of exome sequencing, leads to identification of Novel genes in 10-30% of mutation negative RAS/MAPK syndromes [31,32].

The cardiac hypertrophic response implicates signal transduction pathways initiated by ligand-stimulated membrane-bound receptors (RTKs, GPCRs (G protein-coupled receptors)) and biomechanical stress sensors (integrins). GPCR receptors, also known as seven-transmembrane receptors (7-TM receptors), are a large and diverse family of cell surface receptors found in eukaryotes and play a crucial role in cell signaling [33]. It acts

through EPAC (Exchange Protein Directly Activated by cAMP) and ERK, activate RAS proteins and induce the release of internal Ca^{2+} stores through calcineurin/NFAT (Nuclear Factor of Activated T-cells) proteins to elicit pathological hypertrophy. All these pathways converge on the modulation of transcriptional factors (MEF2 (Myocyte Enhancer Factor 2), JUN and GATA4), which induce the expression of genes of the hypertrophic program. Some “red flags” such as facial dysmorphism, lentigines, sensorineural deafness, PVS (Pulmonary Valvular Stenosis) and biventricular hypertrophy which is reflected by an extreme right axis deviation (a “superior” QRS axis) on the electrocardiogram and represents a specific disease marker [34-37]. Germline mutations in RAF1 are found in 3–5% of NS subjects and show normal valvuloseptal growth, but exhibit eccentric cardiac hypertrophy, probably due to enhanced MEK-ERK signaling [38]. In contrast to non-syndromic primary HCM, HCM in RASopathies confers a high risk of mortality in infancy, which could be attributed to earlier age at presentation and occurrence of heart failure [39]. Among the neurodevelopmental anomalies, the most common is chiari malformation and other findings include delayed myelination, ventriculomegaly, nonspecific white matter abnormalities and corpus callosum hypoplasia [40].

Prenatal Diagnosis of Rasopathies

Prenatal testing should be done when any US finding suggestive of lymphatic dysplasia, CH (cystic hygroma), increased NT (nuchal thickness or translucency) or NF ((Nuchal fold- an ultrasound finding during the second trimester of pregnancy, indicating a thickened area at the back of the fetal neck ($\geq 6\text{mm}$) and it is a soft marker for Down syndrome (Trisomy 21) and other chromosomal abnormalities, though the baby may not have a genetic condition)), hydrops fetalis (HF) or effusions, congenital heart disease (CHD), such as valvular dysplasia or hypertrophic cardiomyopathy (HCM); polyhydramnios (excess amniotic fluid, often $>90\%$), renal anomalies and skeletal abnormalities like short long bones (humerus and femur) and ulnar deviation of wrist [41].

NT (nuchal thickness or translucency) is the amount of fluid behind the fetus's neck in the first trimester of pregnancy detected by ultrasound. A small amount of fluid is normal, and measuring the amount of fluid can help to find the chances of the fetus has a chromosomal or genetic variant. An anechoic space is visible and measurable sonographically in all fetuses between the 11th and the 14th week of the pregnancy as in Figure 1. For most pregnancies, NT above 3 millimeters prompts a discussion of genetic counseling and additional testing. The diagnostic yield of increased NT was significantly higher as 20% when it was greater than 6 mm and 10 % when it was lower than 6 mm and much lower as 1% when this finding was seen as isolated.

The association between the increased NT and the chromosomal abnormalities has been well documented (Figure 2). It helps us to identify the high-risk fetuses for trisomy 21 and other chromosomal abnormalities [42]. The rate of chromosomal aberration and pathogenic CNVs (copy number variations) on chromosomal microarray is high among fetuses with NT between 3.0 and 3.4 mm [43].

Underlying pathophysiological mechanisms for nuchal fluid collection under the skin include cardiac dysfunction, venous

congestion in the head and neck, altered composition of the extracellular matrix, failure of lymphatic drainage, fetal anemia or hypoproteinemia and congenital infection [45]. The abnormal accumulation of nuchal fluid decreases after the 14th week.



Figure 1: Normal Nuchal Translucency Thickness (NT)



Figure 2: Increased Nuchal Translucency Thickness (NT) [44].

Hydrops fetalis as shown in Figure 3 is characterized by abnormal fluid accumulation in fetuses, presents a significant risk of still-birth and neonatal mortality, categorized into immune hydrops due to blood type (RhD)-incompatible pregnancy and NIHF (Non-immune hydrops fetalis) and recent studies have highlighted genetic factors as crucial determinants. Whole-exome sequencing (WES) reported an overall diagnostic rate of 29% to 37% and the predominant disease category observed was RASopathies in NIHF cases affects approximately one in 1700–3000 pregnancies. Functional analysis of the mutant channels revealed a loss-of-function defect in the voltage-dependent sodium channels (Nav), with one channel exhibiting no conduction and the other showing a reduced channel opening due to missense variants of the SCN4A gene encoding these channels [46-49].

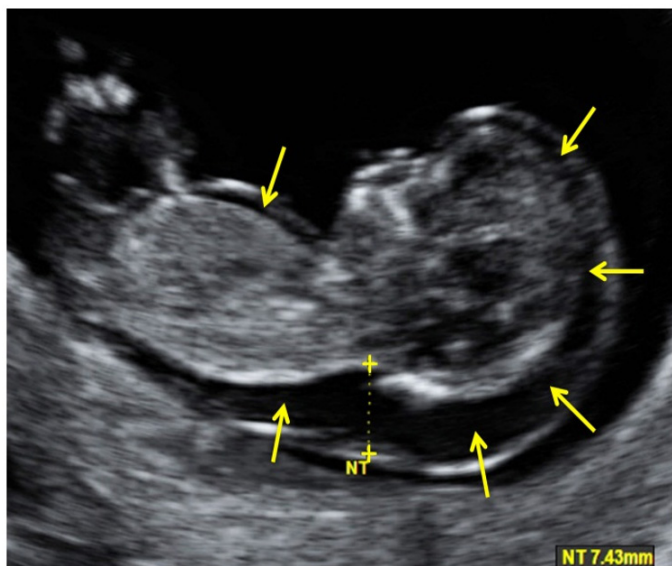


Figure 3: Hydrops Fetalis

The most frequent isolated finding in RASopathy patients is CH (cystic hygroma- 4%), a benign congenital malformation of the lymphatic system due to lack of development of communication between the lymphatic and venous systems and its incidence is approximately 1/6000 live births [50,51]. At the end of the fifth week of gestation, the lymphatic system starts to develop and fluid accumulation within and along the lymphatic vessel tracts and enlargement of the jugular lymphatic sacs are due to this connection failure and may resolve if an alternate route of drainage develops. 70–80% of cystic hygromas occur in the neck, usually in the posterior cervical triangle and 20–30% occurs in the axilla, superior mediastinum, chest wall, mesentery, retro-peritoneal region, pelvis and lower limbs [52,53]. Lymphangiomas may be divided histologically into two major groups based on the depth and the size of abnormal lymph vessels. The superficial ones are called lymphangioma circumscriptum. The more deep seated ones are cavernous lymphangioma or cystic hygroma in areas of areola or loose connective tissues [54]. They are multilocular cysts filled with clear or yellow lymph fluid and brilliantly transilluminant [55]. The mass increases in size and may compresses the neurovascular and respiratory structures as shown in Figure 4 to cause fetal bradycardia and airway obstruction. Noonan syndrome is one of the RASopathies and this group of disorders is often associated with cystic hygroma. Prenatal diagnosis by ultrasound is based on the demonstration of a septated or non-septated, bilateral cystical lesion in the fetal occipitocervical region in both a sagittal and axial plane. Implementation of assessment of the fetal nuchal translucency (NT) in first-trimester screening programs for aneuploidy, the rate of detection has been increased and fetuses with a nuchal translucency thickness more than 10mm and hydropic fetuses had a worse outcome [56].

Prenatal detection of hypertrophic cardiomyopathy (HCM) through advanced echocardiographic imaging is crucial for early diagnosis [58] and its key features include interventricular septal hypertrophy as shown in Figure 5, which serves as a primary diagnostic marker [59].



Figure 4: Cystic Hygroma in the Neck of a Full-Term Infant [57].

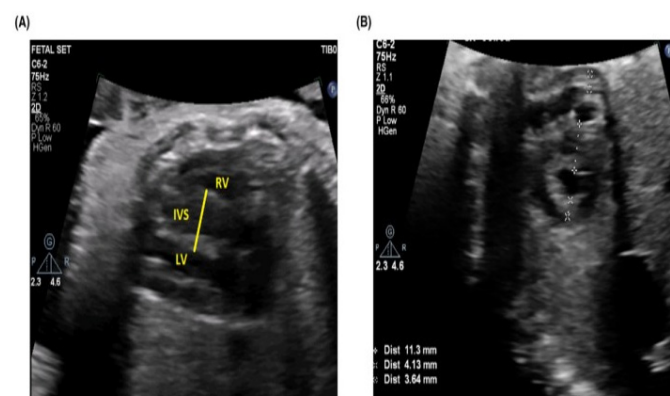


Figure 5: (A) Axial view of the fetal heart demonstrates significant hypertrophy of the interventricular septum (IVS) and cardiomegaly, with clear visualization of the right ventricle (RV) and left ventricle (LV). (B) Short-axis view of both ventricles reveals marked asymmetric hypertrophy of the interventricular septum [60].

Embryonal RMS (Rhabdomyosarcomas) is characterized by a loss of heterozygosity at 11p15.5 and whole or partial gains of chromosomes 2, 8, 12, 13, and/or 20.2 Mutations in the FGFR4/RAS/AKT pathway and increased PTEN (Phosphatase and TENsin homolog) hypermethylation has also been detected in this subtype. Embryonal Rhabdomyosarcoma of the Bladder in a male toddler is shown in Figure 6. The incidence of RMS in patients with CS is 8.7% due to HRAS mutation All individuals with malignancy had a codon 12 mutation (especially G12A) and the rare G12V mutation is associated with a more severe, early lethal phenotype; some patients die from respiratory distress, hypertrophic cardiomyopathy, or malignant tachycardia prior to being diagnosed with CS [61,62].

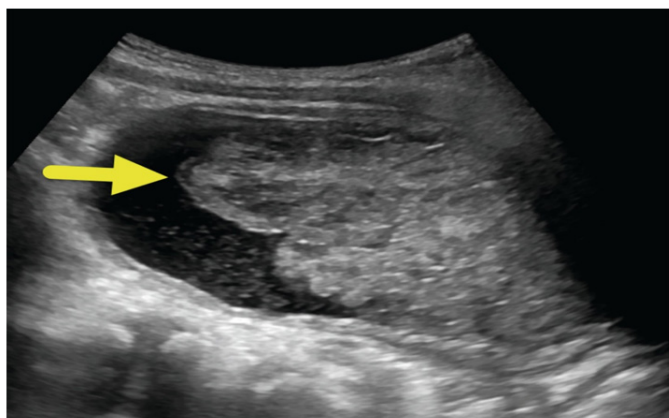


Figure 6: Longitudinal Ultrasound of the Bladder Showing a Lobulated Mass [63].

Operative image indicating the presence of tumor originating in the left medial UL (umbilical ligament) as shown in Figure 7 in a 4.4-year-old female on preterm delivery at 34 weeks and right medial umbilical ligament as shown in Figure 8 in 5.4-year-old female with pregnancy complicated by polyhydramnios.

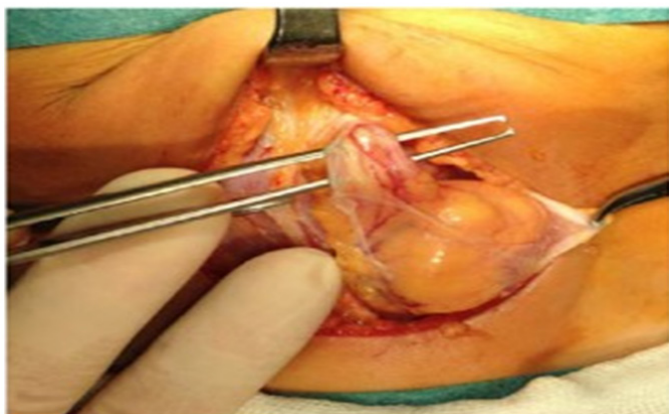


Figure 7: Showing the Rhabdomyosarcoma of Left Medial Umbilical Ligament



Figure 8: Showing the Rhabdomyosarcoma of Right Medial Umbilical Ligament [64].

Historically, NF1 (neurofibromatosis type 1) was the first described RASopathy, diagnosed through clinical analysis and characteristic phenotypical features such as café-au-lait spots,

intertriginous freckling, neurofibromas, and skeletal dysplasia [65]. Genomic DNA analysis of the RASopathy genes to find the pathogenic genetic variants by Sanger sequencing as well as whole-genome sequencing (WGS) and in the case of NS, where approximately 10–20% tested were negative [66]. The second most common type of biomarker used in RASopathies is at the RNA level, involving qualitative and quantitative analysis of mRNA and non-coding RNA (ncRNA), such as microRNAs (miRNAs) and long non-coding RNAs (lncRNAs)[67]. Elevated anti-thyroid peroxidase (anti-TPO) antibody levels (> 500 units/ml) are indeed associated with both Noonan syndrome (NS) and cardiofaciocutaneous syndrome (CFCS), which are RASopathies, found in 59% of patients with thyroiditis but in none of the controls or the patients with non-thyroidal illness [68,69].

The most common cutaneous malignant tumor to have a mutation in the RAS/MAPK pathway is malignant melanoma. A recent study done by Scott et al, used parallel or Sanger sequencing for identifying various mutated genes in prenatal diagnosis. Next-generation sequencing (NGS) has found a few other genes that were altered in patients of RASopathies, but functionally they have been not validated yet. KAT6B, RREB1, and CDC42 alterations have been found to be associated with NS-like features, while a mutation in YWHAZ has been associated with cardio-facio-cutaneous syndrome (CFCS) [70].

Genetic testing in cases of unexplained aborted or sudden cardiac deaths, even in previously healthy children, can be valuable in establishing a diagnosis, determining the prognosis, and assessing risk to family members. A previously healthy infant who suffered aborted sudden cardiac death was found to have a de novo genetic mutation in the SOS1 gene, typical of Noonan syndrome. Ventricular fibrillation arrest associated with a RASopathy in the absence of the typical structural cardiac phenotypes of hypertrophic cardiomyopathy or pulmonary stenosis [71,72]. Genetic mutations that disrupt ion channel function, which are associated with inherited cardiac arrhythmias. Dysfunctional HCN4 (hyperpolarisation-activated cyclic nucleotide-gated) channels might directly cause rhythm disorders related to mutation of SCN5A (9) gene [73].

Therapeutic Options

MEK Inhibitors

The efficacy of small molecule inhibitors for the treatment of HCM associated with the RASopathies, employed knockin strategies and prenatal or postnatal treatment of the MEK inhibitor, PD0325901, successfully rescued embryonic lethality, growth, and cardiac defects in NS-associated *Sos1*E846K mice [74]. Interestingly, although prenatal treatment of the NS-associated *Kras*V14I mutant mice with PD0325901 rescued HCM, when administered postnatally, HCM was not rescued [75]. NS-associated mutant might engage distinct pathways postnatally for the progression of HCM and so treating pregnant mothers with MEK inhibitors, the failure of PD0325901 to reverse HCM postnatally is disappointing. Postnatal treatment of the NS-associated *Raf1*L613V mutant mice with the MEK inhibitor completely rescued HCM with accompanying improvement in cardiac functionality [76].

Trametinib, a highly selective reversible allosteric inhibitor of MEK1/2 activity, was found to reverse progressive HCM in an NS infant harboring either RITS35T or RITF82L mutation. Upon

trametinib treatment, an amelioration of the increased ventricular mass was observed, accompanied by reduced outflow tract obstruction and improved parameters of heart failure [77]. NS can be associated with severe cardiovascular and lymphatic anomalies, potentially lethal during infancy, neonatal and fetal periods and trametinib is a promising drug in these critically ill children [78].

Certain RASopathy mutants will not be responsive to MEK inhibition. The phosphatidylinositol 3'-kinase (PI3K)-AKT pathway positively regulates cardiac tissue mass and AKT activation through the mammalian target of rapamycin (mTOR) increases protein synthesis and prevents muscle atrophy and apoptosis, resulting in cardiomyocyte hypertrophy. Pharmacological studies showed that postnatal treatment of NSML-associated Pptn11Y279C/+ mice with rapamycin, a mTOR inhibitor, prevented the development of HCM and associated increase in cardiomyocyte hypertrophy due to increased AKT activity [79]. PTPN11Q510E mutation that represents a NSML RASopathy who exhibited severe HCM was treated with a mTOR inhibitor, everolimus improved heart failure and decreased the levels of the HCM marker, brain natriuretic peptide. Unlike in the Ptpn11Y279C/+ mice, everolimus treatment in this patient did not reverse the HCM, which could imply that earlier intervention prior to irreversible cardiac remodeling would have been more effective [80].

Src Family Tyrosine Kinase Pathway Inhibition by prenatal and postnatal treatment with low-dose dasatinib in mice also successfully rescued the HCM phenotype [81].

C-Type Natriuretic Peptide Analogues

Achondroplasia is caused by a gain-of-function mutation in the fibroblast growth factor receptor 3 (FGFR3) gene, a member of the tyrosine kinase family, resulting in prolonged activation of RAS/MAPK and alteration of chondrocyte proliferation and differentiation at the growth plate level. An analogue of CNP that is resistant to proteolytic degradation (BMN111/vosoritide; BioMarin) significantly improved growth plate abnormalities and bone growth by reducing RAS/MAPK activation [82].

Statins

Statins were first investigated in the treatment of cognitive impairment in NF1 and lovastatin was shown to improve synaptic plasticity as well as attention and memory in NF1 patients, but not with simvastatin [83-85]. Although they are contraindicated during pregnancy, the clinical effects of statins in pregnant women through an interactive review by the analysis of fifteen original articles within five years demonstrate that statins have not been associated with the development of fetal malformations and it is useful in preventing unfavorable cardiovascular outcomes, with the potential to reduce oxidative stress and angiogenic dysfunction [86]. However, the use of statins to prevent pre-eclampsia in humans has not been properly clarified. Statins may be safe when used during pregnancy since there was no association with congenital anomalies, but caution is needed because of an increased risk of low birth weight and preterm labor [87,88].

An important issue is the optimal age for starting treatment. In several animal studies, MEK (Mitogen-Activated Protein Kinase) inhibitor treatment was initiated during embryonic devel-

opment (by exposing the pregnant mothers to the MEK inhibitor) and continued after birth. MEK inhibition prevented the various developmental defects (i.e. craniofacial, cardiac, and growth defects) when started prenatally, whereas this treatment did not ameliorate these defects when started after weaning [89]. Another issue is the duration of treatment and therapy aimed at allowing normal growth and cognitive development should probably be maintained throughout childhood.

Gene Editing

In utero gene editing has the potential to treat genetic diseases prenatally. Clustered regularly interspaced short palindromic repeats (CRISPR)-CRISPR-associated 9 (CRISPR-Cas9) or base editor 3 (BE3) in utero, seeking therapeutic modification of Pcsk9 or Hpd in wild-type mice or the murine model [90]. Intronic CRISPR repair is a therapeutic strategy to treat NS (Noonan Syndrome)-associated hypertrophic cardiomyopathy [91]. A CRISPR-based epigenetic repression system downregulating MAP2K1 and MAP2K2 expression for the treatment of RASopathies was proposed in animal models [92].

Discussion

In utero manifestations of RASopathies are less well characterized and approximately half of RASopathy variants being inherited in postnatal series [93]. The Noonan syndrome has been well established in its association with NIHF (Nonimmune hydrops fetalis), defined by the presence of fetal ascites, pleural or pericardial effusions, skin edema, cystic hygroma, increased thickness of nuchal translucency (≥ 3.5 mm), or a combination of these conditions. Pregnant women with fetuses that have NIHF are also at risk for complications resulting from a form of pre-eclampsia called mirror syndrome [94,95]. Some genetic disorders underlying NIHF portend mild long-term outcomes, whereas others are lethal despite treatment [96,97]. Increased index of suspicion of Costello syndrome (CS) in newborn is shown in Table 2.

Table 2: Features to Suspect CS in Newborn

1.	Fetal atrial tachycardia
2.	Increased birth weight and head circumference
3.	Neonatal hypoglycemia
4.	severe feeding difficulties
5.	Urinalysis for hematuria – embryonal rhabdomyosarcoma
6.	Loose, redundant skin on the hands and feet seen in newborns – key role in clinical suspicion of CS

Prenatal diagnosis of HCM is critical, as it enables the clinicians to anticipate potential complications such as LVOTO (Left ventricular outflow tract obstruction), arrhythmias, and heart failure, which may manifest shortly after birth [98]. Interventricular septal thickness measured during fetal life is a reliable predictor of postnatal outcomes, with a threshold of ≥ 4.5 mm is associated with higher risk of perinatal morbidity and mortality [99]. Long-term outcomes of fetal HCM are influenced by underlying etiology. Parasternal long axis view showing “ventriculomegaly” (massive RVH and LVH) as shown in Figure 9 is well tolerated in a 9-year-old boy with Costello syndrome as in Figures 10 and 11 with increased RV pressure as in Figure 12

due to increased capillary growth along with the hypertrophied muscle [100-102].

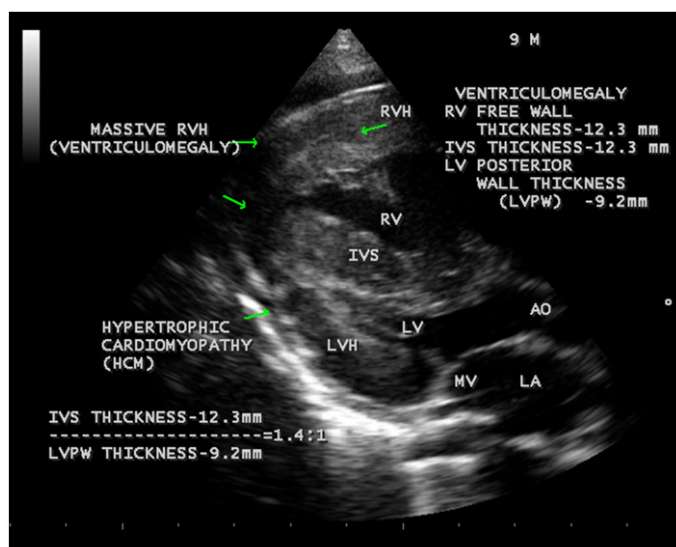


Figure 9: Showing Biventricular Hypertrophy (Ventriculomegaly) in a 9-Year-Old Boy with Costello Syndrome



Figure 11: Tight Tendoachilles and Plantar Hyperkeratosis in a 9-Year-Old Boy with Costello Syndrome



Figure 10: Radiant Smile, Warm, Sociable Personality and Skin Folding below the Lower eyelids of Costello Syndrome in a 9-Year-Old Boy

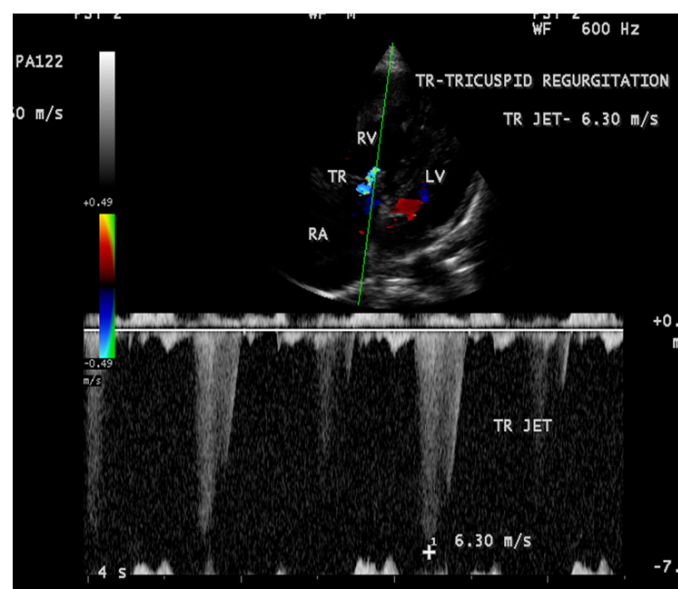


Figure 12: Apical 4 Chamber View – CW Doppler showing High Velocity TR (Tricuspid Regurgitation) Jet with Early Peaking Suggesting High RV Pressure at Suprasystemic Levels-165 mmHg

Conclusion

Rasopathy testing is recommended when the fetus shows an isolated increased NT ≥ 5.0 mm or when NT of ≥ 3.5 mm and at least one of the following ultrasound anomalies is present: distended JLS (Jugular Lymphatic Sacs), hydrops fetalis, polyhydramnios, pleural effusion, ascites, cardiac defects and renal anomalies [103]. Invasive prenatal testing is proposed for fetuses with NT values above 2.5 mm. Rapid screening for chromosomal aneuploidy (13, 18, 21, X, and Y), chromosomal microarray (CMA), and sequencing of the genes involved in Noonan syndrome and RASopathies are routinely performed. Whole genome sequencing identified two compound heterozygous variants in the NUP107 gene in fetuses [104]. Rigosertib, a novel dual Ras/MAPK and PI3K/AKT pathway inhibitor, reverses Hypertrophic

Cardiomyopathy in RAF1-Associated Noonan Syndrome [105]. The MEK inhibitor trametinib was an effective drug therapy for cardiac hypertrophy in NS mice with heterozygous LZTR1 (Leucine-zipper-like posttranslational regulator 1) mutation [106]. More than half of the RASopathy cases in utero manifests during the first trimester and a cutoff value of NT $\geq$ 6 mm missed 45% of cases during this period. Exome sequencing is a preferable option due to its efficiency within the limited timeframe in prenatal settings [107].

References

1. Tidyman WE, Rauken KA (2016) Expansion of the RASopathies. *Current Genetic Medicine Reports* 4: 57-64.
2. Aoki Y, Niihori T, Inoue S, Matsubara Y (2016) Recent advances in RASopathies *Journal of Human Genetics* 61: 33-39.
3. Gripp KW, Lisa Schill, Lisa Schoyer, Beth Stronach, Anton M Bennett, et al. (2020) The sixth international RASopathies symposium: precision medicine-From promise to practice. *American Journal of Medical Genetics Part A* 182: 597-606.
4. Scott A, Di Giosaffatte N, Pinna V, Daniele P (2021) When to test fetuses for RASopathies? Proposition from a systematic analysis of 352 multicenter cases and a postnatal cohort. *Genetics in Medicine* 6: 1116-1124.
5. Leach NT, Wilson Mathews DR, Rosenblum LS, Zhou Z, Zhu H, et al. (2019) Comparative assessment of gene-specific variant distribution in prenatal and postnatal cohorts tested for Noonan syndrome and related conditions. *Genetics in Medicine* 21: 417-425.
6. Grande M, Jansen FAR, Blumenfeld YJ, A Fisher, A O Odibo, et al. (2015) Genomic microarray in fetuses with increased nuchal translucency and normal karyotype: a systematic review and meta-analysis. *Ultrasound in Obstetrics & Gynecology* 46: 650-658.
7. Stark Z, Lunke S, Brett GR, Tan NB, Stapleton R, et al. (2018) Meeting the challenges of implementing rapid genomic testing in acute pediatric care. *Genetics in Medicine* 12: 1554-1563.
8. Petrovski S, Aggarwal V, Giordano JL (2019) Whole-exome sequencing in the evaluation of fetal structural anomalies: a prospective cohort study. *Lancet* 393: 758-767.
9. Gabriel H, Korinith D, Ritthaler M (2021) Trio exome sequencing is highly relevant in prenatal diagnostics. *Prenatal Diagnosis* 1-7.
10. Teresa N, Billie R, Rebecca R (2020) Exome Sequencing for Prenatal Diagnosis in Nonimmune Hydrops Fetalis. *New England Journal of Medicine* 18: 1746-1756.
11. Zhang W, Liu H (2002) MAPK signal pathways in the regulation of cell proliferation in mammalian cells. *Cell Research* 12: 9-18.
12. Farcas A, Janosi L (2024) GTP-Bound N-Ras Conformational States and Substates Are Modulated by Membrane and Point Mutation. *International Journal of Molecular Sciences* 25: 1430.
13. Ali S, Ali U, Qamar A, Zafar I, Yaqoob M, et al. (2023) Predicting the effects of rare genetic variants on oncogenic signaling pathways: A computational analysis of HRAS protein function. *Frontiers in Chemistry* 11: 1173624.
14. Sarah Lee (2025) Understanding the role of stop codons in protein synthesis and their impact on genetic disorders. *Ultimate Guide to Stop Codons in Genetics*.
15. Steff S, Nishi H, Petukh M, Panchenko AR, Alexov E (2013) Molecular mechanisms of disease-causing missense mutations. *Journal of molecular biology* 425: 3919-3936.
16. Woodward AA, Urbanowicz RJ, Naj AC, Moore JH (2022) Genetic heterogeneity: Challenges, impacts, and methods through an associative lens. *Genetic Epidemiology* 46: 555-571.
17. Mohiuddin M, Kooy RF, Pearson CE (2022) De novo mutations, genetic mosaicism and human disease. *Frontiers in Genetics* 13: 983668.
18. Pudewell Silke, Wittich Christoph, Kazemein Jasemi Neda, Bazgir Farhad, Ahmadian Reza (2021) Accessory proteins of the RAS-MAPK pathway: moving from the side line to the front line. *Communications Biology* 4: 10.
19. Parada LF, Tabin CJ, Shih C (1982) Bladder carcinoma oncogene is homologue of Harvey sarcoma virus ras gene. *Nature* 297: 474-478.
20. Gripp KW, Morse LA, Axelrad M, Chatfield KC, Chidekel A, et al. (2019) Costello syndrome: Clinical phenotype, genotype, and management guidelines. *American Journal of Medical Genetics* 179: 1725-1744.
21. Gripp KW, Lin AE, Stabley DL (2006) HRAS Mutation Analysis in Costello Syndrome: Genotype and Phenotype Correlation. *American Journal of Medical Genetics* 140: 1-7.
22. Gripp KW, Lin AE (2012) Costello syndrome: a Ras/mitogen activated protein kinase pathway syndrome (rasopathy) resulting from HRAS germline mutations. *Genetics in Medicine* 14: 285-292.
23. Weaver KN, Wang D, Cnota J, Gardner N (2014) Early-lethal Costello syndrome due to rare HRAS Tandem Base substitution (c.35_36GC>AA; p.G12E)-associated pulmonary vascular disease. *Pediatrics And Developmental Pathology* 17: 421-430.
24. Burkitt-Wright EM, Bradley L, Shorto J, McConnell VP (2012) Neonatal lethal Costello syndrome and unusual dinucleotide deletion/insertion mutations in HRAS predicting p.Gly12Val. *American Journal of Medical Genetics* 158: 1102-1110.
25. Bertola D, Buscarilli M, Stabley DL (2017) Phenotypic spectrum of Costello syndrome individuals harboring the rare HRAS mutation p.Gly13Asp. *American Journal of Medical Genetics* 173: 1309-1318.
26. Stiazaran-Symonds, E, Ney GM, Higgs C, Oba L (2023) Cancer in Costello syndrome: a systematic review and meta-analysis. *British Journal of Cancer* 128: 2089-2096.
27. Bhopal Mohapatra, Gulzar Ahmad, Scott Nadeau (2013) Protein tyrosine kinase regulation by ubiquitination: Critical roles of Cbl-family ubiquitin ligases. *Biochimica et Biophysica Acta (BBA) - Molecular Cell Research* 1833: 122-139.
28. Jafari D, Mousavi MJ, Keshavarz Shahbaz S, Jafarzadeh L (2021) E3 ubiquitin ligase Casitas B lineage lymphoma-b and its potential therapeutic implications for immunotherapy. *Clinical and Experimental Immunology* 204: 14-31.
29. Ren J, Lv L, Tao X, Zhai X, Chen X (2024) The role of CBL family ubiquitin ligases in cancer progression and therapeutic strategies. *Frontiers in Pharmacology* 26: 1432545.
30. Yoshida T, Muramatsu H, Wakamatsu M, Sajiki D (2023) Clinical and molecular features of CBL-mutated juvenile myelomonocytic leukemia. *Haematologica* 108: 3115-3119.

31. Aoki Y, Matsubara Y (2013) Ras/MAPK syndromes and childhood hemato-oncological diseases. *International Journal of Hematology* 97: 30-36.
32. Lip Hen Moey, Go Hun Seo, Boon Eu Cheah, Wee Teik Keng, Hane Lee, et al. (2025) A first large study of whole-exome sequencing (WES) in 489 patients with suspected rare genetic disorders at a tertiary centre in Malaysia. *Rare* 3: 100102.
33. Sala V, Gallo S, Leo C (2012) Signaling to Cardiac Hypertrophy: Insights from Human and Mouse RASopathies. *Molecular Medicine* 18: 938-947.
34. Chen L, Song M, Yao C (2021) Calcineurin in development and disease. *Genes & Diseases* 9: 915-927.
35. Razzaq SS, Khan I, Naeem N, Salim A, Begum S, et al. (2022) Overexpression of GATA binding protein 4 and myocyte enhancer factor 2C induces differentiation of mesenchymal stem cells into cardiac-like cells. *World Journal of Stem Cells* 14: 700-713.
36. Limongelli G, Monda E, Tramonte S, Gragnano F, Masarone D, et al. (2020) Prevalence and clinical significance of red flags in patients with hypertrophic cardiomyopathy. *International Journal of Cardiology* 299: 186-191.
37. Rapezzi C, Arbustini E, Caforio AL, Charron P (2013) Diagnostic work-up in cardiomyopathies: Bridging the gap between clinical phenotypes and final diagnosis. A position statement from the ESC Working Group on Myocardial and Pericardial Diseases. *European Heart Journal* 34: 1448-1458.
38. Wu X, Simpson J, Hong JH, Kim KH, Thavarajah NK, et al. (2011) MEK-ERK pathway modulation ameliorates disease phenotypes in a mouse model of Noonan syndrome associated with the Raf1(L613V) mutation. *Journal of Clinical Investigation* 121: 1009-1025.
39. Norrish G, Kolt G, Cervi E, Field E, Dady K, et al. (2021) Clinical presentation and long-term outcomes of infantile hypertrophic cardiomyopathy: A European multicentre study. *ESC Heart Failure* 8: 5057-5067.
40. Reynolds G, Gazzin A, Carli D, Massuras S, Cardaropoli S, et al. (2025) Update on the Clinical and Molecular Characterization of Noonan Syndrome and Other RASopathies: A Retrospective Study and Systematic Review. *International Journal of Molecular Sciences* 26: 3515.
41. Stuurman KE, Marieke Joosten, Ineke van der Burgt, Mariet Elting, Helger G Yntema, et al. (2019) Prenatal ultrasound findings of rasopathies in a cohort of 424 fetuses: update on genetic testing in the NGS era1. *Journal of Medical Genetics* 56: 654-661.
42. Nicolaides KH, Brizot ML, Snijders RJM (1994) Fetal nuchal translucency: ultrasound screening for fetal trisomy in the first trimester of pregnancy. *BJOG: An International Journal of Obstetrics & Gynaecology* 101: 782-786.
43. Vriendt MDE, Rooryck C, Madar H, Coatleven F, Vincienne M, et al. (2025) Outcomes associated with fetal nuchal translucency between 3.0 and 3.4mm in the first trimester, *Acta Obstetrica et Gynecologica Scandinavica* 104: 629-636.
44. Gersak K, Darija Strah DM, Pohar-Perme M (2013) Increased Fetal Nuchal Translucency Thickness and Normal Karyotype: Prenatal and Postnatal Outcome, Down Syndrome. InTech.
45. Nicolaides KH (2004) The 11-13+6 weeks scan. London: Fetal Medicine Foundation.
46. Sparks TN, Lianoglou BR, Adami RR, Pluym ID, Holliman K, et al. (2020) Exome sequencing for prenatal diagnosis in nonimmune hydrops fetalis. *New England Journal of Medicine* 383: 1746-1756.
47. Mone F, Eberhardt RY, Hurles ME, McMullan DJ, Maher ER, et al. (2021) Fetal hydrops and the Incremental yield of Next-generation sequencing over standard prenatal Diagnostic testing (FIND) study: prospective cohort study and meta-analysis. *Ultrasound in Obstetrics & Gynecology* 58: 509-518.
48. Al-Kouatly HB, Shivashankar K, Mossayebi MH, Makhameh M, Critchlow E, et al. (2023) Diagnostic yield from prenatal exome sequencing for non-immune hydrops fetalis: A systematic review and meta-analysis. *Clinical Genetics* 103: 503-512.
49. Kubota T, Nagata M, Takagi K (2025) Hydrops fetalis due to loss of function of hNav1.4 channel via compound heterozygous variants, *Journal of Human Genetics* 70: 3-8.
50. Scott A, Di Giosaffatte N, Pinna V (2021) When to test fetuses for RASopathies? Proposition from a systematic analysis of 352 multicenter cases and a postnatal cohort. *Genetics in Medicine* 23: 1116-1124.
51. Dahnert W (1993) Radiology review manual. 2. Arizona: Williams & Wilkins.
52. Song TB, Kim CH, Kim SM, Kim YH, Byun JS, et al. (2002) Fetal axillary cystic hygroma detected by prenatal ultrasonography: a case report. *Journal of Korean Medical Science* 17: 400-402.
53. Manikoth P, Mangalore GP, Megha V (2004) Axillary Cystic Hygroma. *Journal of Postgraduate Medicine* 50: 215-216.
54. Mordehai J, Kurzbart E, Shinhar D, Sagi A, Finaly R, et al. (1998) Lymphangioma circumscriptum. *Pediatric Surgery International* 13: 208-210.
55. Rasidaki M, Sifakis S, Vardaki E, Koumantakis E (2005) Prenatal diagnosis of a fetal chest wall cystic lymphangioma using ultrasonography and MRI: a case report with literature review. *Fetal Diagnosis and Therapy* 20: 504-507.
56. Lore Schreurs L, Lore Lannoo L, De Catte L, Van Schoubroeck D, Devriendt K, et al. (2018) First trimester cystic hygroma colli: Retrospective analysis in a tertiary center. *European Journal of Obstetrics & Gynecology and Reproductive Biology* 231: 60-64.
57. Sannoh S, Quezada E, Merer DM, Moscatello A, Golombek SG (2009) Cystic hygroma and potential airway obstruction in a newborn: a case report and review of the literature. *Cases Journal* 2: 48.
58. Gasior T (2024) Advances in Cardiac Imaging and Genetic Testing for Diagnosis and Risk Stratification in Cardiomyopathies: 2024 Update. *Journal of Clinical Medicine* 13: 7166.
59. Mandeş L, Roşca M, Ciupercă D, Popescu B A (2020) The Role of Echocardiography for Diagnosis and Prognostic Stratification in Hypertrophic Cardiomyopathy. *Journal of Echocardiography* 18: 137-148.
60. Golbabaei A, Moghaddam ESA, Naemi M, Mohammad Talebi H (2025) Hypertrophic Cardiomyopathy in a Fetus: A Rare Presentation Case Report. *Clinical Case Reports* 13: e70599.
61. Gripp KW, Lin AE (2012) Costello syndrome: a Ras/mitogen activated protein kinase pathway syndrome (rasopathy)

- resulting from HRAS germline mutations. *Genetics in Medicine* 14: 285-292.
62. Quezada E, Gripp KW (2007) Costello syndrome and related disorders. *Current Opinion in Pediatrics* 19: 636-644.
 63. Ruoyan Zhu, Richard B Towbin, Carrie M Schaefer, Alexander J Towbin (2023) Embryonal Rhabdomyosarcoma of the Bladder. *Applied Radiology* 6: 40-42.
 64. Sánchez-Montenegro C, Vilanova-Sánchez A, Barrena-Delfa S, Tenorio J (2017) Costello Syndrome and Umbilical Ligament Rhabdomyosarcoma in Two Pediatric Patients: Case Reports and Review of the Literature. *Case Reports in Genetics* 1587610.
 65. Tidyman WE, Rauen KA (2009) The RASopathies: Developmental Syndromes of Ras/MAPK Pathway Dysregulation, *Current Opinion in Genetics and Development*. 19: 230-236.
 66. Zenker M (2022) Clinical Overview on RASopathies. *American Journal of Medical Genetics Part C: Seminars in Medical Genetics* 190: 414-424.
 67. Amani V, Riemondy KA (2023) Integration of Single-Nuclei RNA-Sequencing, Spatial Transcriptomics and Histochemistry Defines the Complex Microenvironment of NF1-Associated Plexiform Neurofibromas. *Acta Neuropathologica Communications* 11: 158.
 68. Siano MA, Pivonello R, Salerno M, Falco M, Mauro C, et al. (2022) Endocrine System Involvement in Patients with RASopathies: A Case Series. *Frontiers in Endocrinology* 13: 1030398.
 69. Engler H, Riesen WF, Keller B (1994) Anti-thyroid peroxidase (anti-TPO) antibodies in thyroid diseases, non-thyroidal illness and controls. Clinical validity of a new commercial method for detection of anti-TPO (thyroid microsomal) autoantibodies, *Clinica Chimica Acta* 225: 123-136.
 70. Padhiyar Jigna, Mahajan Rahul, Panda Maitreyee (2024) Evolving Concepts in Pathogenetics, Clinical Features, and Management. *Indian Dermatology Online Journal* 15: 392-404.
 71. Follansbee CW, Malloy-Walton L (2021) Ventricular fibrillation due to a likely pathogenic SOS1 variant: An unrecognized etiology of infantile sudden death?. *HeartRhythm Case Reports* 7: 510-513.
 72. Cirelli Michael, Wackel Philip, Javed Rabia, Gavrilova Ralitz, Qureshi Muhammad, et al. (2024) SOS1-Related Noonan Syndrome and Sudden Cardiac Arrest in the Absence of Cardiomyopathy-An Arrhythmia Phenotype?. *American journal of medical genetics. Part A* 197: e63912.
 73. Liu S, Wang W, Yang Y, Huang Z (2025) Ventricular ion channels and arrhythmias: an overview of physiology, pathophysiology and pharmacology, *Medical Review* 5: 231-243.
 74. Chen PC, Wakimoto H, Conner D, Araki T, Yuan T, et al. (2010) Activation of multiple signaling pathways causes developmental defects in mice with a Noonan syndrome-associated Sos1 mutation. *Journal of Clinical Investigation* 120: 4353-4365.
 75. Hernandez-Porras I, Fabbiano S, Schuhmacher AJ, Aicher A, Canamero M, et al. (2014) K-RasV14I recapitulates Noonan syndrome in mice. *Proceedings of the National Academy of Sciences of the United States of America* 111: 16395-16400.
 76. Wu X, Simpson J, Hong JH, Kim KH, Thavarajah N, et al. (2011) MEK-ERK pathway modulation ameliorates disease phenotypes in a mouse model of Noonan syndrome associated with the Raf 1(L613V) mutation. *Journal of Clinical Investigation* 121: 1009-1025.
 77. Andelfinger G, Marquis C, Raboisson MJ, Theoret Y, Waldmuller S, et al. (2019) Hypertrophic cardiomyopathy in Noonan syndrome treated by MEK-inhibition. *Journal of American College of Cardiology* 73: 2237-2239.
 78. Isabel DB, Juan L, Léon B, Arthur S, An VD, et al. (2025) Trametinib as a targeted treatment in cardiac and lymphatic presentations of Noonan syndrome. *Frontiers in Pediatrics* 13: 1475143.
 79. Marin TM, Keith K, Davies B, Conner DA, Guha P, et al. (2011) Rapamycin reverses hypertrophic cardiomyopathy in a mouse model of LEOPARD syndrome-associated PTPN11 mutation. *Journal of Clinical Investigation* 121: 1026-1043.
 80. Hahn A, Lauriol J, Thul J, Behnke-Hall K, Logeswaran T, et al. (2015) Rapidly progressive hypertrophic cardiomyopathy in an infant with Noonan syndrome with multiple lentigines: palliative treatment with a rapamycin analog. *American Journal of Medical Genetics* 167: 744-751.
 81. Yi JS, Huang Y, Kwaczala AT, Kuo IY, Ehrlich BE, et al. (2016) Low-dose dasatinib rescues cardiac function in Noonan syndrome. *JCI Insight* 20: e90220.
 82. Lorget F, Kaci N, Peng J, Benoist-Lasselin C, Mugniery E, et al. (2012) Evaluation of the therapeutic potential of a CNP analog in a Fgfr3 mouse model recapitulating achondroplasia. *American Journal of Human Genetics* 91: 1108-1114
 83. Acosta MT, Kardel PG, Walsh KS, Rosenbaum KN, Gioia GA, et al. (2011) Lovastatin as treatment for neurocognitive deficits in neurofibromatosis type 1: phase I study. *Pediatric Neurology* 45: 241-245.
 84. Mainberger F, Jung NH, Zenker M, Wahllander U, Freudenberg L, et al. (2013) Lovastatin improves impaired synaptic plasticity and phasic alertness in patients with neurofibromatosis type 1. *BMC Neurology* 13: 131
 85. Van der Vaart T, Plasschaert E, Rietman AB, Renard M, Oostenbrink R, et al. (2013) Simvastatin for cognitive deficits and behavioural problems in patients with neurofibromatosis type 1 (NF1-SIMCODA): a randomised, placebo-controlled trial, *The Lancet Neurology* 12: 1076-1083.
 86. Lins Serafim J, Lucas Santos de Menezes Teles P, Souza Lima AK, Dos Santos Coelho, J, Luna Maranhão Conrado P, et al. (2025) Clinical repercussions of statin use during pregnancy: a review of the literature. *Brazilian Journal of Gynecology and Obstetrics (RBGO)* 47: e-rbgo2.
 87. Chang J, Chen Y, Chen I, Lin W, Chen Y, et al. (2021) Perinatal Outcomes After Statin Exposure During Pregnancy. *JAMA Network Open* 4: e2141321.
 88. Poornima IG, Pulipati VP, Brinton EA, Wild RA (2023) Update on Statin Use in Pregnancy. *American Journal of Medicine* 136: 12-14.
 89. Hernandez-Porras I, Fabbiano S, Schuhmacher AJ, Aicher A, Canamero M, et al. (2014) K-RasV14I recapitulates Noonan syndrome in mice. *Proceedings of the National Academy of Sciences of the United States of America* 111: 16395-16400.

90. Rossidis AC, Stratigis JD, Chadwick AC (2018) In utero CRISPR-mediated therapeutic editing of metabolic genes. *Nature Medicine* 24: 1513-1518.
91. Hanses Ulrich, Kleinsorge M, Lennart Roos, Gökhan Yigit, Yun Li, et al. (2020) Intronic CRISPR Repair in a Preclinical Model of Noonan Syndrome-Associated Cardiomyopathy. *Circulation* 142: 1059-1076.
92. WO2024238906 (2024) Epigenetic Repression Protein Therapy for Rasopathies.
93. Gelb BD, Cavé H, Dillon MW (2018) ClinGen's RASopathy Expert Panel consensus methods for variant interpretation. *Genetics in Medicine* 20: 1334-1345.
94. Gedikbasi A, Oztarhan K, Gunenc Z (2011) Preeclampsia due to fetal non-immune hydrops: mirror syndrome and review of literature. *Hypertension in Pregnancy* 30: 322-330.
95. Society for Maternal-Fetal Medicine (SMFM) (2015) Clinical guideline #7: nonimmune hydrops fetalis. *American Journal of Obstetrics & Gynecology* 212: 127-139.
96. Derderian SC, Trivedi S, Farrell J (2014) Outcomes of fetal intervention for primary hydrothorax. *Journal of Pediatric Surgery* 49: 900-903.
97. Mardy AH, Chetty SP, Norton ME, Sparks TN (2019) A system-based approach to the genetic etiologies of non-immune hydrops fetalis. *Prenatal Diagnosis* 39: 732-750.
98. Wolf CM (2019) Hypertrophic Cardiomyopathy: Genetics and Clinical Perspectives. *Cardiovascular Diagnosis and Therapy* 9: S388-S415.
99. Elmekawi SF, Mansour GM, Elsafty MS, Hassanin AS, Laban M, et al. (2015) Prediction of Fetal Hypertrophic Cardiomyopathy in Diabetic Pregnancies Compared With Postnatal Outcome. *Clinical Medicine Insights: Women's Health* 8: 39-43.
100. Ramachandran Muthiah (2022) Ventriculomegaly in Costello syndrome. 11th International Conference on Cardiology and Cardiovascular Medicine Research, Current Research: Cardiology 9.
101. Muthiah R (2016) Costello Syndrome with Congenital Pulmonary Valve Stenosis and Ventriculomegaly: A Case Report. *Case Reports in Clinical Medicine* 5: 250-267.
102. Rudolph AM (1974) *Congenital Diseases of the Heart*. Yearbook Medical Chicago.
103. Stuurman KE, Joosten M, Van der Burgt I, Elting M, Yntema HG, et al. (2019) Prenatal ultrasound findings of rasopathies in a cohort of 424 fetuses: update on genetic testing in the NGS era. *Journal of Medical Genetics* 56: 654-661.
104. Atallah I, Cisarova K, Guenot C, Dubruc E, Superti-Furga A, et al. (2025) Recurrent Increased Nuchal Translucency Led to the Identification of Novel NUP107 Variants. *American Journal of Medical Genetics* 197: e63920
105. Legler L, Xu B (2024) Rigosertib Reverses Hypertrophic Cardiomyopathy in RAF1-Associated Noonan Syndrome. *bioRxiv* 11: 623865.
106. Abe T, Takada S, Aoki Y (2024) Dysregulation of RAS proteostasis by autosomal-dominant LZTR1 mutation induces Noonan syndrome-like phenotypes in mice. *JCI Insight* 9: e182382.
107. Qiu-Xia Yu, Li Zhen, Yong-Ling Zhang, Si-Yun Li, Ru Li, Dong-Zhi Li, (2025) Clinical and molecular analysis of seventy-one fetal cases with RASopathies, *European Journal of Obstetrics & Gynecology and Reproductive Biology*, 314, 114691.